OPINION Open Access

# Immune responses in the irritable bowel syndromes: time to consider the small intestine

Grace L. Burns<sup>1,2,3</sup>, Nicholas J. Talley<sup>1,2,3</sup> and Simon Keely<sup>1,2,3</sup>\*

# **Abstract**

**Background:** Irritable bowel syndrome (IBS) is considered a disorder of gut-brain interaction (DGBI), presenting as chronic abdominal pain and altered defaecation. Symptoms are often food related. Much work in the field has focused on identifying physiological, immune and microbial abnormalities in the colon of patients; however, evidence of small intestinal immune activation and microbial imbalance has been reported in small studies. The significance of such findings has been largely underappreciated despite a growing body of work implicating small intestinal homeostatic imbalance in the pathogenesis of DGBIs.

**Main text:** Small intestinal mechanosensation is a characteristic feature of IBS. Furthermore, altered small intestinal barrier functions have been demonstrated in IBS patients with the diarrhoea-predominant subtype. Small intestinal bacterial overgrowth and increased populations of small intestinal mast cells are frequently associated with IBS, implicating microbial imbalance and low-grade inflammation in the pathogenesis of IBS. Furthermore, reports of localised food hypersensitivity responses in IBS patients implicate the small intestine as the site of immune-microbial-food interactions.

**Conclusions:** Given the association of IBS symptoms with food intake in a large proportion of patients and the emerging evidence of immune activation in these patients, the current literature suggests the pathogenesis of IBS is not limited to the colon but rather may involve dysfunction of the entire intestinal tract. It remains unclear if regional variation in IBS pathology explains the various symptom phenotypes and further work should consider the intestinal tract as a whole to answer this question.

**Keywords:** Irritable bowel syndrome, Disorders of gut-brain interaction, Functional gastrointestinal disorders, Small intestine, Immune

# **Background**

Irritable bowel syndrome (IBS) is a disorder of gutbrain interaction (DGBI), the term for heterogeneous conditions of the gastrointestinal tract (GIT), previously described as 'functional' gastrointestinal disorders, for which there is no recognised overt structural pathology [1]. Instead, IBS is diagnosed based on a specific symptom profile, including abdominal pain

in conjunction with alterations in bowel habit. IBS patients are subtyped based on bowel habit profile, into diarrhoea (IBS-D), constipation (IBS-C), mixed (IBS-M) or indeterminate, and it is suggested that approximately 10–20% of all cases develop after an episode of acute gastroenteritis (post-infectious, PI-IBS) [2]. Recent findings of subtle, sub-clinical gastrointestinal inflammatory changes in these patients indicate a role for the immune system in driving symptom onset and chronicity [3]; however, the nature of immune activation and involvement remains unknown. The absence of obvious pathology or a known trigger of immune involvement limits the therapeutic options and

Full list of author information is available at the end of the article



<sup>\*</sup>Correspondence: simon.keely@newcastle.edu.au

<sup>&</sup>lt;sup>2</sup> College of Health, Medicine and Wellbeing, The University of Newcastle, Callaghan. New South Wales. Australia

Burns et al. BMC Medicine (2022) 20:115 Page 2 of 9

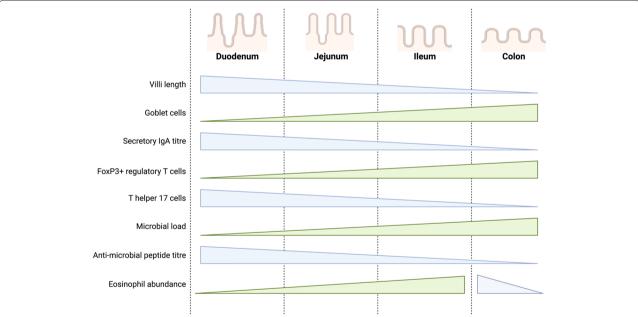
diagnostic approaches for these patients to often ineffective management of symptoms rather than specific treatment of the cause. It is hypothesised that IBS represents several conditions that result from dysfunction of the pathways that regulate homeostasis [4]; however, this is yet to be conclusively proven.

To date, most studies investigating the immune abnormalities in IBS have focused on the colon, given the association of the condition with altered defaecation patterns and technical difficulties in sampling biopsies or fluid from the small intestine (SI), particularly in larger studies. However, the SI is the major site of both nutrient antigen exposure and maintenance of immune tolerance against food and commensal microbes [5]. Combining these facts with emerging evidence for food intolerance as a driver of IBS [6, 7], the SI is increasingly being explored as a site of IBS pathology [8–10]. As such, we review the literature for SI involvement in immune responses in IBS and advocate for a more universal approach to examining immune activation across the diverse geography of the gut in future studies.

# Main text

# Regional specificity and homeostasis along the gastrointestinal tract

The GIT exhibits regional specificity, best demonstrated by differences in the structure and function of the SI and colon. The primary function of the SI is the absorption of nutrients, with most absorption occurring in the duodenum and jejunum due to the increased surface area provided by the villi structures characteristic of these sites [11]. Colonic function, in contrast, is associated with the absorption of water and processing indigestible food material into faeces for elimination [11]. These site-specific functions are associated with physiological changes throughout the length of the GIT, with the SI exhibiting a lower pH [12] and shorter transit time than the colon [13]. In addition, there are site-specific selective pressures on the microbiota due to such physiological characteristics of the SI and colon [14]. Consequently, the immune profiles of the proximal and distal GIT are associated with geographical luminal signals (Fig. 1), such as exposure to dietary antigens in the SI and microbial signals in the colon [15], although much of this work



**Fig. 1** Regional specificity of selected immune and microbial components in the gastrointestinal tract. There is a distinct variation in the abundance of immune and microbial factors that mediate homeostasis in conjunction with physiological function throughout the small intestine and colon. Because of the role of the small intestine in nutrient absorption, the proximal segments (duodenum and jejunum) have longer, finger-like villi to increase the available surface area. The distal small intestine, the ileum, has shorter villi. Functionally, the colon primarily reabsorbs water and processes unabsorbable waste as faeces for elimination and does not have the finger-like projections of the small intestine. In the colon, immune homeostasis is primarily focused on tolerating the high commensal burden. As such, there is a higher abundance of Th type 17 cells in the duodenum that decreases towards the distal colon, corresponding with an inverse abundance of FoxP3<sup>+</sup> regulatory T cells. Nutrient absorption capacity is greatest in the duodenum and decreases towards the colon. This corresponds with the small intestinal immune systems focus on oral tolerance to food antigens and production of anti-microbial peptide production and secretory IgA. Eosinophils are a normal constituent of the gastrointestinal tract. Their abundance increases towards the distal small intestine, peaking in the terminal ileum and proximal colon before decreasing towards the rectum [15–18]. The image was created using BioRender.com

Burns et al. BMC Medicine (2022) 20:115 Page 3 of 9

has been demonstrated in animal models, rather than in humans.

In the SI, the immune system modulates homeostasis against luminal antigens by an active process known as oral tolerance [19] in conjunction with the small intestinal mucus layer, which facilitates closer contact of commensals with epithelial cells for sampling by antigenpresenting cells [20]. The discrimination of detrimental antigens from innocuous food proteins and commensals within the GIT is a complex process mediated by the actions of either reactive lymphocytes or regulatory T cells (Treg). Oral tolerance exists in order to prevent redundant and excessive immune responses to common food antigens and intestinal flora [21]. Ingested food proteins undergo a rigorous digestive process before reaching the small intestine, including digestion by proteases in the saliva, stomach and pancreatic acids [22]. Despite this digestive process, some proteins retain intact epitope structures that may come into contact with the mucosa of the lumen [23]. It is at this point that the immune system directs the development of tolerance against the specific epitope encountered to prevent unnecessary immune responses; however, in situations of homeostatic interruption, tolerance may be replaced by the induction of adaptive hypersensitivity immune responses.

# Involvement of the colonic adaptive immune system in IBS

While there is little consensus regarding the specific immune profile of IBS in the literature, largely due to the innate heterogeneity in the condition itself and methodological differences across studies [4], IBS patients seem to have greater basal levels of immune activation compared to outpatient or healthy control populations. A meta-analysis of cytokine studies [24] found an imbalance in the ratio of pro-inflammatory tumour necrosis factor (TNF) to interleukin (IL-)10. IL-10 is considered anti-inflammatory due to its capacity to limit T cell differentiation to prevent T helper (Th) cell polarisation [25], highlighting homeostatic imbalance as a feature of IBS. A small study of peripheral monocytes stimulated with lipopolysaccharide suggested monocytes from IBS patients were more mature [26] and IBS patients have higher levels of T cells expressing markers of activation compared to controls [27, 28]. Findings of altered Tolllike receptor (TLR) expression in the colon [29, 30] and elevated faecal antimicrobial  $\beta$ -defensin 2 levels [31] suggest activation of the innate immune system by microbial components may also contribute to disease pathogenesis, and this is supported by an exaggerated release of inflammatory cytokines (IL-1β, IL-6, IL-8 and TNF) from whole blood samples stimulated with TLR agonists in patients compared to healthy controls [32]. Importantly a metaanalysis of colonic immune cells highlighted regional and subtype-specific differences in immune cell numbers [33], supporting the notion of IBS as a condition not localised to one region of the colon.

The most consistently reported feature of IBS is increased mast cell numbers in both the SI and colon [34–40]. While some studies were unable to demonstrate altered mast cell numbers in the colon [41] likely due to methodological differences, sample sizes or selection bias, increased secretion of colonic tryptase and histamine [41-43] support a role for mast cells in IBS. Furthermore, the proximity of mast cells to enteric nerves correlates with the severity of abdominal pain, visceral hypersensitivity, fatigue and co-morbid depression [37, 44, 45] in IBS, suggesting a prominent role for these cells in both the pathophysiology and psychological burden of IBS. Similarly, colonic eosinophils have been reported as increased in patients [46-49]; however, this finding is not reproduced in all studies [50]. While there is a paucity of IBS studies examining the eosinophil number and activation status in the small intestine, one study found no change in duodenal eosinophil number in IBS patients compared to controls [35]. Eosinophils have gained prominence as an effector cell in other DGBIs including functional dyspepsia (FD), and given their relationship with mast cells in sensitisation-like immune responses [51], it is likely there is a functional role for eosinophils in SI immune activation in IBS. However, currently, the signals recruiting and activating these cells are unknown.

Given the finding of increased mast cells in IBS, a prominent hypothesis for immune activation is the notion of antigens, likely of food and/or microbial origins, stimulating the induction of a Th type 2 response. In this setting, antigens are presented to naïve T cells by antigen-presenting cells (such as dendritic cells) which drive differentiation into activated Th2 cells to stimulate immunoglobulin (Ig) E production from B cells. The subsequent binding of IgE to mast cells and re-exposure to antigen then results in degranulation and release of inflammatory mediators in close proximity to nerve cells that result in the onset of symptoms [52]. While one study demonstrated that stimulation of peripheral T cells from IBS patients resulted in increased production of IL-5 and IL-13 [53], a systematic review of the literature revealed there was little specific evidence for activation of this pathway in IBS [4].

The potential for Th17 responses in the microinflammatory profile of IBS has also been proposed, based on indirect evidence of increased peripheral TNF and IL-6 in patients [3, 54]. Th17 cells exist in a balance with Tregs to maintain gut immune homeostasis [55]; however, Th17 responses can also induce inflammation and autoimmune responses. For instance, in asthma, activation of Th17 pathways results in the release of IL-17, a cytokine which

Burns et al. BMC Medicine (2022) 20:115 Page 4 of 9

acts on the epithelium to drive recruitment of effector cells, including macrophages and eosinophils [56]. Interestingly, one study showed serum levels of IL-17a and TNF were significantly increased in conjunction with decreased IL-10 levels in patients with IBS-D [57], implicating an altered Th17/Treg axis in this subtype. While a meta-analysis of colonic immune cells in IBS found the total lymphocyte population (CD3<sup>+</sup>) was increased in patients, likely due to increased CD4<sup>+</sup> cells [33], there are few studies examining the intestinal or colonic T cell phenotypes to support the Th2/Th17 hypotheses. Given both Th2 and Th17 immune responses would likely be occurring in relation to luminal antigens and the duodenum is where antigens initially interact with the immune system, the notion of IBS as a condition exclusively affecting the colon does not make sense. Rather, it is likely that IBS represents an adaptive immune response to luminal antigens that manifests heterogeneously along both the SI and colon.

# Evidence for small intestinal immune involvement in IBS

Reduced integrity of the mucosal barrier in IBS likely facilitates translocation of luminal antigens for direct contact with the immune system [58], which cyclically promotes continued permeability of the barrier. Increased SI [59, 60] and colonic barrier permeability have been associated with visceral hypersensitivity [59], independent of disease subtype [61], suggesting that a loss of barrier integrity may be the first step to priming of the immune system in IBS. However, one study identified SI permeability was attributable to the IBS-D subtype only, finding that altered SI permeability in IBS-C compared to controls was influenced by confounding lifestyle factors and that colonic permeability was unchanged when measured using multi-sugar testing [62]. Interestingly, dysregulated stress responses may mediate immune activation in the SI in IBS, given the association between corticotropin-releasing factor and jejunal mast cells and eosinophils [63, 64]. Such dysregulation has also been demonstrated in FD [65], and it remains to be seen if this pathway occurs in conjunction with or independently of the classical Th2-mediated response initially proposed to drive immune activation in IBS.

While the specific contribution of T cell populations to the presentation of IBS remains unclear, studies have suggested that IBS patients have a greater lymphocyte burden in the duodenum [35, 66] and jejunum [67], and there is no change in the total lymphocyte density in the ileum [45, 68]. Early work using the first iteration of the Rome criteria [67] identified subclinical increases in intraepithelial lymphocytes (IEL) and infiltration of lymphocytes into the jejunal myenteric plexus in IBS patients when compared to outpatient controls. The increase in

IEL number has been reported in further studies, specifically in IBS-D [36, 66] and in the terminal ileum [69], and suggests enhanced surveillance of luminal content or lingering hyperreactivity of the SI immune system. Given the role of the myenteric plexus in the coordination of contraction and motility [70], lymphocytic infiltration at this site suggests subclinical inflammation specific to the enteric system that may be linked to motility dysfunction in patients. In addition, qualitative assessment of jejunal mast cells found no difference between patients and controls [67], a finding that was later supported by a systematic review and meta-analysis of mast cells in the SI [40], which identified increases in ileal, but not duodenal or jejunal mast cells in IBS patients. One of the only population studies to examine SI pathology in IBS identified increased IELs (specific to IBS-C only) and mast cells in the duodenum of both diarrhoea and constipation subtypes [35]. While the literature is conflicting regarding the role of lamina propria lymphocytes in the SI in IBS, likely due to variation in methodology for quantification and patient categorisation, it does appear that alteration in immune cell populations is a SI feature of a subset of patients. Interestingly, increased lamina propria lymphocyte populations were not reported in the ileum in two studies [45, 68], while the meta-analysis [40] suggested that increased SI mast cells are a feature specifically of the ileum. Unfortunately, progress towards confirming and understanding an immune mechanism to drive symptom chronicity in IBS will depend on the identification of specific antigenic triggers, and given the heterogeneity among patients, this process will be complicated given it is likely that no single antigen is responsible for IBS symptoms.

# Food antigens as a trigger for IBS symptom onset

Up to 84% of IBS patients self-report that food ingestion induces symptoms, with incompletely absorbed carbohydrate sources (such as dairy, beans and some fruits) and foods that drive histamine release (including milk, beer and pork) most reported with symptoms [71]. Furthermore, the exclusion of foods with raised IgG titres in a trial of 150 patients resulted in a significant symptom reduction at 12 weeks [72], linking food-driven responses to symptom burden. The introduction of specific foods directly to the duodenum of IBS patients using confocal laser endomicroscopy (CLE) showed that 70% of these patients had a detectable response (CLE+) to one or more foods [7]. Furthermore, there were significant differences in the immune activation profile of CLE+ and CLE— IBS patients, characterised by increases in IEL counts, increased claudin-2 and decreased occuludin levels, suggestive of barrier dysfunction in CLE+ patients. This response profile was not associated with systemic Burns et al. BMC Medicine (2022) 20:115 Page 5 of 9

IgE; however, CLE+ IBS patients had higher levels of eosinophil degranulation [7], suggestive of a non-IgEmediated food intolerance. The findings of this study confirm the capacity for a SI immune response to food in a subset of susceptible IBS patients. However, it is unclear how prevalent this phenomenon is in IBS patients, and there are likely other mechanisms by which IBS symptoms may manifest. Such differences in manifestations of IBS would help to explain the heterogeneity reported regarding immune activation profiles in the literature [4]. A more recent study demonstrated localised responses to injection of food antigens in the recto-sigmoid region of IBS patients, characterised by oedema, IgE antibody production and mast cell activation at the challenge site [6]. While intact food proteins are unlikely to make direct contact with the rectosigmoid mucosa during the process of digestion, this study demonstrates immune responses to food are localised to the intestinal mucosa in IBS. Here, loss of oral tolerance to common food antigens may result from heightened immunosurveillance and drive visceral hypersensitivity [6]. When considered with studies using CLE to examine responses to antigen [7], these findings would suggest that both classical (IgE-mediated) and non-classical hypersensitivity pathways may be activated in IBS patients. Importantly, the identification of these localised immune responses with no systemic profile suggests the need for caution when interpreting studies of systemic mediators (e.g. serum cytokines or peripheral blood mononuclear cell populations) in IBS patients.

# Small intestinal bacterial overgrowth (SIBO) and the microbiota in IBS

The colonic and faecal microbiomes have been profiled in IBS patients and suggest altered composition is a feature of IBS [73-76]. The findings of a systematic review suggested specific phylums (Proteobacteria and Bacteroidetes) and families (Enterobacteriaceae and Lactobacillaceae) of bacteria likely contributed to a proinflammatory microenvironment in the colon in IBS [73]. Furthermore, a systematic review of faecal microbiota transplantation (FMT) for IBS found that administration of the transplant to the small intestine was effective, and the placebo effect for this route of administration was lower compared to FMT administered via colonoscopy [77]. Such data suggests targeting of the SI microbiota may be an effective approach to treating IBS, and findings of altered anti-microbial defence factors (such as TLRs and  $\beta$ -defensin 2, as previously discussed) in patients highlight dysregulation of the immune response to commensals may be associated with immune activation. Increased production of IgA in the terminal ileum may be a consequence of shifts in the total microbial composition [78], resulting in local inflammatory signals. However, the lack of consistent microbial profiling methods and patient characterisation combined with a paucity of functional data regarding the altered microbes limits our understanding of the specific species and taxa that may contribute to IBS. Furthermore, there are very little data regarding the SI luminal or mucosa-associated microbiome outside of the context of SIBO.

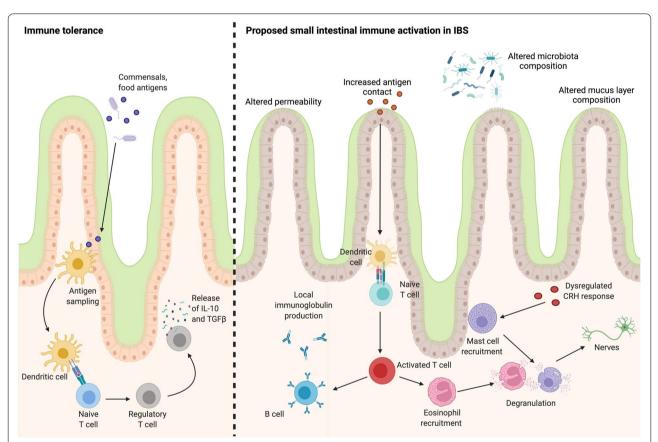
SIBO describes excessive overgrowth of colonic-type bacteria, classically defined as > 10<sup>5</sup> colony-forming units per millilitre of upper gastrointestinal aspirate [79] although a cut-off of 10<sup>3</sup> has been suggested more recently [80]. SIBO has been associated with IBS, although the literature is conflicting regarding whether the association is specific to one subtype over another [81-83]. The frequency of SIBO in IBS patients has been reported in the range of 4-78% [84]; however, there is substantial variation in the methodologies used to quantify bacterial load and a lack of consensus diagnostic criteria that have hampered efforts to firmly investigate the relationship between SIBO and IBS to date [85]. While breath testing has become the preferred diagnostic choice given the non-invasiveness and simplicity of such approaches [86], it is of note this method is influenced by gut transit and patient factors including physical activity [87] and pausing medications such as proton pump inhibitors before the test [88] and has poor correlation with gold standard aspirate cultures [89]. As such, it is difficult to determine if the abnormal test results reflect the presence of SIBO, or more rapid transit and fermentation by colonic bacteria [89]. There is a suggestion that the expansion of a colonic-like microbial profile in the small intestine triggers the immune system to induce the low-grade inflammatory state and induces hypersensitivity responses. In this scenario, the overgrowth of particular species stimulates immune cells to secrete proinflammatory cytokines, including IL-1 $\alpha$  and IL-1 $\beta$  [90], that drive recruitment of effector cells, such as mast cells, and impair the mucosal barrier. Furthermore, treatment with oral rifaximin, a broad-spectrum antibiotic that acts in the SI, has been shown to relieve symptoms of abdominal bloating and pain in IBS-D patients [91]. Animal studies suggest the efficacy of rifaximin may be due to its capacity to downregulate inflammatory cytokines including IL-17, IL-6 and TNF while improving intestinal barrier permeability and reducing visceral hypersensitivity [92]. These findings highlight that not only is the SI involved in IBS but targeting of microinflammation at this site may result in improved symptom burden.

Significant progress has been made in characterising IBS as a condition of disordered interactions between the gut and the brain with microinflammation as a central pathology. This concept has directly challenged the

Burns et al. BMC Medicine (2022) 20:115 Page 6 of 9

notion of this condition as a 'functional' disorder, given the significant array of physiological, microbial and immune abnormalities described. However, it is time to progress the field further towards greater consideration of the role of the SI in this condition. While this is not a new concept, given early reports of altered SI permeability in IBS patients, the SI has largely been ignored in the search for targetable mechanisms that underlie GI dysfunction in IBS. The literature suggests that homeostatic imbalance is not limited to the colon, given that the immune system, microbiota and physiological function of the SI are affected by IBS. A proposed pathway for small intestinal immune activation in IBS is included

in Fig. 2. What is unclear is if the involvement of the SI is a feature of a subset of patients, common to all or is instead a previously unrecognised link between IBS and other DGBIs, such as functional dyspepsia. FD affects the gastroduodenal region, with altered barrier function and microinflammation described in the duodenum [93]. Like IBS, food and microbial antigens are hypothesised as responsible for cyclic episodes of symptom onset due to subclinical inflammation. Interestingly, studies suggest between 26.7 and 48.7% of IBS patients meet the Rome criteria for concurrent FD [94]. In addition, this patient subset reports greater symptom severity and decreased quality of life compared to patients with only one DGBI



**Fig. 2** Hypothesised immune mechanisms potentially involved in small intestinal dysfunction in IBS. The small intestinal immune system actively modulates tolerance to commensal microbes and food components to maintain homeostasis, in conjunction with the mucosal barrier and mucus layer. In this process, antigens are sampled by dendritic cells and presented to naïve T cells. The differentiation of these cells to regulatory T cells results in the release of interleukin-10 and transforming growth factor beta, which actively suppresses inflammatory immune responses. In contrast, physiological abnormalities in the composition of the mucus layer, coupled with altered mucosal permeability and changed microbial community composition in IBS patients may allow for increased antigen contact with the mucosa and a dysregulated or increased stress response. In this environment, antigen presentation may result in the activation of T cell subsets that drive B cell maturation and specific antibody production that is likely localised to the gastrointestinal tract. The activation of the adaptive immune system may drive the recruitment of eosinophils and mast cells, which degranulate and release inflammatory mediators. The release of these mediators near enteric nerves is likely to promote abnormal signalling and may result in visceral pain. Altered or enhanced stress signalling may also enhance this eosinophil and mast cell response to further contribute to immune activation. However, these pathways require further investigation in IBS patients compared to controls to demonstrate the mechanisms underlying intestinal immune activation. The image was created using BioRender.com

Burns et al. BMC Medicine (2022) 20:115 Page 7 of 9

[95], leading to questions of whether these are distinct conditions or rather different manifestations of the same process of homeostatic imbalance and microinflammation. In support of this, one hypothesis suggests that the site of gastroenteritis predicts the development of post-infectious DBGIs [96, 97], whereby infections in the proximal SI are more likely to result in FD development, while distal infections may predispose to onset of IBS and if both regions are involved, then overlapping FD/IBS may develop. However, currently, there is little prospective data to support this concept.

While the literature regarding the phenotype of T cell activation in IBS is underdeveloped, it is worth considering that significant alterations in individual effector T cell populations are unlikely in IBS. Rather, future studies should consider the deep characterisation of the effector and memory T cell populations in both the SI and colon. If these are in fact disorders of homeostatic imbalance, it is likely that the T cell repertoire is instead characterised by shifts in the balance of regulatory, effector and memory T cells.

### **Conclusions**

Despite reports of SI alterations in physiology, microbial communities and immune activation in IBS patients, many studies continue to focus solely on the colon. However, dysfunction of the entire intestinal tract may be implicated in IBS, and this will be an important consideration in future studies as we move towards identifying specific triggers and immune pathways that drive symptom chronicity. Characterisation of both SI and colonic immune profiles in large cohorts will be critical to unravelling the heterogeneity inherent to IBS and may eventually identify distinct subgroups of people based on responses to food and/microbial luminal antigens, allowing for specific therapeutic targeting.

# Abbreviations

CD: Cluster of differentiation; CLE: Confocal laser endomicroscopy; DGBI: Disorder of gut-brain interaction; FD: Functional dyspepsia; GIT: Gastrointestinal tract; IBS: Irritable bowel syndrome; IBS-C: Irritable bowel syndrome constipation; IBS-D: Irritable bowel syndrome diarrhoea; IBS-M: Irritable bowel syndrome mixed; IEL: Intraepithelial lymphocytes; Ig: Immunoglobulin; IL: Interleukin; PI-IBS: Post-infectious irritable bowel syndrome; SI: Small intestine; SIBO: Small intestinal bacterial overgrowth; Th: T helper; TLR: Toll-like receptor; TNF: Tumour necrosis factor; Treg: Regulatory T cells.

# Acknowledgements

The authors acknowledge the funding by the National Health and Medical Research Council (NHMRC) for the Centre of Research Excellence in Digestive Health.

# Authors' contributions

GLB wrote the original draft of the manuscript. NJT and SK reviewed and edited the manuscript. Resources were provided by NJT and SK. All authors read and approved the final manuscript.

#### **Funding**

National Health and Medical Research Council (NHMRC) for the Centre of Research Excellence in Digestive Health.

# Availability of data and materials

N/A

# **Declarations**

# Ethics approval and consent to participate

N/A

## Consent for publication

All authors provide consent for publication.

# **Competing interests**

GLB declares that she has no competing interests.

NJT: Dr. Talley reports non-financial support from HVN National Science Challenge NZ, personal fees from Aviro Health (Digestive health) (2019), Anatara Life Sciences, Brisbane (2019), Allakos (gastric eosinophilic disease) (2021), Bayer [IBS] (2020), Danone (Probiotic) (2018), Planet Innovation (Gas capsule IBS) (2020), Takeda, Japan (gastroparesis) (2019), twoXAR (2019) (IBS drugs), Viscera Labs, (USA 2021) (IBS-diarrhoea), Dr. Falk Pharma (2020) (EoE), Censa, Wellesley MA USA (2019) (Diabetic gastroparesis), Cadila PharmIncaceuticals (CME) (2019), Progenity Inc. San Diego, (USA 2019) (Intestinal capsule), Sanofiaventis, Sydney (2019) (Probiotic), Glutagen (2020) (Celiac disease), ARENA Pharmaceuticals (2019) (Abdominal pain), IsoThrive (2021) (oesophageal microbiome), BluMaiden (2021), Rose Pharma (2021) and Intrinsic Medicine (2021) outside the submitted work. In addition, Dr. Talley has a patent Nepean Dyspepsia Index (NDI) 1998, Biomarkers of IBS licenced, a patent Licensing Questionnaires Talley Bowel Disease Questionnaire licenced to Mayo/ Talley, a patent Nestec European Patent licenced, and a patent Singapore Provisional Patent "Microbiota Modulation Of BDNF Tissue Repair Pathway" issued. Committees: Australian Medical Council (AMC) [Council Member]; Australian Telehealth Integration Programme; MBS Review Taskforce; NHMRC Principal Committee (Research Committee) Asia Pacific Association of Medical Journal Editors. Boards: GESA Board Member, Sax Institute, Committees of the Presidents of Medical Colleges. Community group: Advisory Board, IFFGD (International Foundation for Functional GI Disorders). Miscellaneous: Avant Foundation (judging of research grants). Editorial: Medical Journal of Australia (Editor in Chief), Up to Date (Section Editor), Precision and Future Medicine, Sungkyunkwan University School of Medicine, South Korea, Med (Journal of Cell Press). Dr. Talley is supported by funding from the National Health and Medical Research Council (NHMRC) to the Centre for Research Excellence in Digestive Health and he holds an NHMRC Investigator grant. SK: Grant/research support: National Health and Medical Research Council (Ideas Grant and Centre for Research Excellence) Viscera Labs (Research contract) and Microba Life Science (research contract). Consultant/advisory boards: Gossamer Bio (Scientific Advisory Board), Anatara Lifescience (Scientific Advisory Board), and Microba Life Science (Consultancy).

# **Author details**

<sup>1</sup>NHMRC Centre of Research Excellence in Digestive Health, The University of Newcastle, Callaghan, New South Wales, Australia. <sup>2</sup>College of Health, Medicine and Wellbeing, The University of Newcastle, Callaghan, New South Wales, Australia. <sup>3</sup>Immune Health Program, Hunter Medical Research Institute, New Lambton Heights, New South Wales, Australia.

Received: 25 October 2021 Accepted: 15 February 2022 Published online: 31 March 2022

# References

- Drossman DA. Functional gastrointestinal disorders: history, pathophysiology, clinical features and Rome IV. Gastroenterology. 2016;150(6):1262–1279.e2.
- Kibune Nagasako C, et al. Irritable bowel syndrome subtypes: clinical and psychological features, body mass index and comorbidities. Rev Esp Enferm Dig. 2016;108(2):59–64.

Burns et al. BMC Medicine (2022) 20:115 Page 8 of 9

- Burns G, et al. Immune activation in functional gastrointestinal disorders. Gastroenterol Hepatol (NY). 2019;15(10):539–48.
- Burns G, et al. Evidence for local and systemic immune activation in functional dyspepsia and the irritable bowel syndrome: a systematic review.
  Am J Gastroenterol. 2019;114(3):429–36.
- Worbs T, et al. Oral tolerance originates in the intestinal immune system and relies on antigen carriage by dendritic cells. J Exp Med. 2006;203(3):519–27.
- Aguilera-Lizarraga J, et al. Local immune response to food antigens drives meal-induced abdominal pain. Nature. 2021;590:151–6.
- Fritscher-Ravens A, et al. Many patients with irritable bowel syndrome have atypical food allergies not associated with immunoglobulin E. Gastroenterology. 2019;157(1):109–118 e5.
- Turcotte J-F, et al. Breaks in the wall: increased gaps in the intestinal epithelium of irritable bowel syndrome patients identified by confocal laser endomicroscopy (with videos). Gastrointest Endosc. 2013;77(4):624–30.
- Grover M, et al. Intestinal chemosensitivity in irritable bowel syndrome associates with small intestinal TRPV channel expression. Aliment Pharmacol Ther. 2021;54(9):1179–92.
- Wang XJ, et al. Differential mRNA expression in ileal mucosal biopsies of patients with diarrhea- or constipation-predominant irritable bowel syndrome. Clin Transl Gastroenterol. 2021;12(4):e00329.
- 11. Mowat AM, Agace WW. Regional specialization within the intestinal immune system. Nat Rev Immunol. 2014;14(10):667–85.
- Fallingborg J. Intraluminal pH of the human gastrointestinal tract. Dan Med Bull. 1999;46(3):183–96.
- Proano M, et al. Transit of solids through the human colon: regional quantification in the unprepared bowel. Am J Physiol. 1990;258(6 Pt 1):G856–62.
- Donaldson GP, Lee SM, Mazmanian SK. Gut biogeography of the bacterial microbiota. Nat Rev Microbiol. 2016;14(1):20–32.
- Agace WW, McCoy KD. Regionalized development and maintenance of the intestinal adaptive immune landscape. Immunity. 2017;46(4):532–48.
- Gebert N, et al. Region-specific proteome changes of the intestinal epithelium during aging and dietary restriction. Cell Rep. 2020;31(4):107565.
- 17. Yantiss RK. Eosinophils in the GI tract: how many is too many and what do they mean? Mod Pathol. 2015;28(S1):S7–S21.
- 18. Straumann A, Simon HU. The physiological and pathophysiological roles of eosinophils in the gastrointestinal tract. Allergy. 2004;59(1):15–25.
- Dubois A, et al. Regulation of Th2 responses and allergic inflammation through bystander activation of CD8+T lymphocytes in early life. J Immunol. 2010;185(2):884–91.
- Parrish A, et al. Intestinal mucus barrier: a missing piece of the puzzle in food allergy. Trends Mol Med. 2022;28(1):36–50.
- Pabst O, Mowat AM. Oral tolerance to food protein. Mucosal Immunol. 2012;5(3):232–9.
- 22. Weiner HL, et al. Oral tolerance. Immunol Rev. 2011;241(1):241–59.
- 23. Vickery BP, et al. Mechanisms of immune tolerance relevant to food allergy. J Allergy Clin Immunol. 2011;127(3):576–84.
- Bashashati M, et al. Cytokine imbalance in irritable bowel syndrome: a systematic review and meta-analysis. Neurogastroenterol Motil. 2014;26(7):1036–48.
- Couper KN, Blount DG, Riley EM. IL-10: the master regulator of immunity to infection. J Immunol. 2008;180(9):5771–7.
- Rodríguez-Fandiño OA, et al. Maturation phenotype of peripheral blood monocyte/macrophage after stimulation with lipopolysaccharides in irritable bowel syndrome. J Neurogastroenterol Motil. 2017;23(2):281–8.
- Nasser Y, et al. Activation of peripheral blood CD4+T-cells in IBS is not associated with gastrointestinal or psychological symptoms. Sci Rep. 2019;9(1):3710.
- Ohman L, et al. T-cell activation in patients with irritable bowel syndrome. Am J Gastroenterol. 2009;104(5):1205–12.
- Belmonte L, et al. Role of Toll like receptors in irritable bowel syndrome: differential mucosal immune activation according to the disease subtype. PLoS One. 2012;7(8):e42777.
- 30. Brint EK, et al. Differential expression of Toll-like receptors in patients with irritable bowel syndrome. Am J Gastroenterol. 2011;106(2):329–36.
- Langhorst J, et al. Elevated human beta-defensin-2 levels indicate an activation of the innate immune system in patients with irritable bowel syndrome. Am J Gastroenterol. 2009;104(2):404–10.

- 32. McKernan DP, et al. Altered peripheral toll-like receptor responses in the irritable bowel syndrome. Aliment Pharmacol Ther. 2011;33(9):1045–52.
- Bashashati M, et al. Colonic immune cells in irritable bowel syndrome: a systematic review and meta-analysis. Neurogastroenterol Motil. 2018;30(1):e13192.
- Vicario M, et al. Increased humoral immunity in the jejunum of diarrhoeapredominant irritable bowel syndrome associated with clinical manifestations. Gut. 2015;64(9):1379–88.
- Walker MM, et al. Duodenal mastocytosis, eosinophilia and intraepithelial lymphocytosis as possible disease markers in the irritable bowel syndrome and functional dyspepsia. Aliment Pharmacol Ther. 2009;29(7):765–73.
- Guilarte M, et al. Diarrhoea-predominant IBS patients show mast cell activation and hyperplasia in the jejunum. Gut. 2007;56(2):203–9.
- Barbara G, et al. Activated mast cells in proximity to colonic nerves correlate with abdominal pain in irritable bowel syndrome. Gastroenterology. 2004;126(3):693–702.
- O'Sullivan M, et al. Increased mast cells in the irritable bowel syndrome. Neurogastroenterol Motil. 2000;12(5):449–57.
- Weston AP, et al. Terminal ileal mucosal mast cells in irritable bowel syndrome. Dig Dis Sci. 1993;38(9):1590–5.
- Robles A, et al. Mast cells are increased in the small intestinal mucosa of patients with irritable bowel syndrome: a systematic review and metaanalysis. Neurogastroenterol Motil. 2019;31(12):e13718.
- 41. Cenac N, et al. Role for protease activity in visceral pain in irritable bowel syndrome. J Clin Investig. 2007;117(3):636–47.
- Barbara G, et al. Mast cell-dependent excitation of visceral-nociceptive sensory neurons in irritable bowel syndrome. Gastroenterology. 2007;132(1):26–37.
- 43. Vivinus-Nébot M, et al. Combination of allergic factors can worsen diarrheic irritable bowel syndrome: role of barrier defects and mast cells. Am J Gastroenterol. 2012;107(1):75–81.
- Piche T, et al. Mast cells and cellularity of the colonic mucosa correlated with fatigue and depression in irritable bowel syndrome. Gut. 2008;57(4):468–73.
- Park JH, et al. Mucosal mast cell counts correlate with visceral hypersensitivity in patients with diarrhea predominant irritable bowel syndrome. J Gastroenterol Hepatol. 2006;21(1 Pt 1):71–8.
- Casado-Bedmar M, et al. Elevated F-EDN correlates with mucosal eosinophil degranulation in patients with IBS—a possible association with microbiota? J Leukoc Biol. 2021.1-11.
- De Silva AP, et al. Subclinical mucosal inflammation in diarrhea-predominant irritable bowel syndrome (IBS) in a tropical setting. Scand J Gastroenterol. 2012;47(6):619–24.
- 48. Singh M, et al. The relationship between mucosal inflammatory cells, specific symptoms, and psychological functioning in youth with irritable bowel syndrome. Sci Rep. 2020;10(1):11988.
- Katinios G, et al. Increased colonic epithelial permeability and mucosal eosinophilia in ulcerative colitis in remission compared with irritable bowel syndrome and health. Inflamm Bowel Dis. 2020;26(7):974–84.
- Willot S, et al. Nerve growth factor content is increased in the rectal mucosa of children with diarrhea-predominant irritable bowel syndrome. Neurogastroenterol Motil. 2012;24(8):734–e347.
- Stone KD, Prussin C, Metcalfe DD. IgE, mast cells, basophils, and eosinophils. J Allergy Clin Immunol. 2010;125(2):S73–80.
- 52. Gurram RK, Zhu J. Orchestration between ILC2s and Th2 cells in shaping type 2 immune responses. Cell Mol Immunol. 2019;16(3):225–35.
- Kindt S, et al. Immune dysfunction in patients with functional gastrointestinal disorders. Neurogastroenterol Motil. 2009;21(4):389–98.
- Liebregts T, et al. Immune activation in patients with irritable bowel syndrome. Gastroenterology. 2007;132(3):913–20.
- Cheng H, et al. The Th17/Treg cell balance: a gut microbiota-modulated story. Microorganisms. 2019;7(12):583.
- Newcomb DC, Peebles RS Jr. Th17-mediated inflammation in asthma. Curr Opin Immunol. 2013;25(6):755–60.
- Choghakhori R, et al. Inflammatory cytokines and oxidative stress biomarkers in irritable bowel syndrome: association with digestive symptoms and quality of life. Cytokine. 2017;93:34–43.
- 58. Turner JR. Intestinal mucosal barrier function in health and disease. Nat Rev Immunol. 2009;9(11):799–809.

Burns et al. BMC Medicine (2022) 20:115 Page 9 of 9

- 59. Zhou Q, Zhang B, Verne NG. Intestinal membrane permeability and hypersensitivity in the irritable bowel syndrome. Pain. 2009;146(1):41–6.
- Martínez C, et al. Diarrhoea-predominant irritable bowel syndrome: an organic disorder with structural abnormalities in the jejunal epithelial barrier. Gut. 2013;62(8):1160–8.
- Camilleri M, et al. Intestinal barrier function in health and gastrointestinal disease. Neurogastroenterol Motil. 2012;24(6):503–12.
- 62. Mujagic Z, et al. Small intestinal permeability is increased in diarrhoea predominant IBS, while alterations in gastroduodenal permeability in all IBS subtypes are largely attributable to confounders. Aliment Pharmacol Ther. 2014;40(3):288–97.
- Salvo-Romero E, et al. Overexpression of corticotropin-releasing factor in intestinal mucosal eosinophils is associated with clinical severity in diarrhea-predominant irritable bowel syndrome. Sci Rep. 2020;10(1):20706.
- 64. Guilarte M, et al. Peripheral corticotropin-releasing factor triggers jejunal mast cell activation and abdominal pain in patients with diarrhea-predominant irritable bowel syndrome. Am J Gastroenterol. 2020;115(12):2047–59.
- Bruce JK, et al. Defects in NLRP6, autophagy and goblet cell homeostasis are associated with reduced duodenal CRH receptor 2 expression in patients with functional dyspepsia. Brain Behav Immun. 2022;101:335–45.
- Foley S, et al. Impaired uptake of serotonin by platelets from patients with irritable bowel syndrome correlates with duodenal immune activation. Gastroenterology. 2011;140(5):1434–43 e1.
- Törnblom H, et al. Full-thickness biopsy of the jejunum reveals inflammation and enteric neuropathy in irritable bowel syndrome. Gastroenterology. 2002;123(6):1972–9.
- Park JH, et al. Enteroendocrine cell counts correlate with visceral hypersensitivity in patients with diarrhoea-predominant irritable bowel syndrome. Neurogastroenterol Motil. 2006;18(7):539–46.
- Talley NJ, et al. Ileocolonic histopathological and microbial alterations in the irritable bowel syndrome: a nested community case-control study. Clin Transl Gastroenterol. 2020;12(1):e00296.
- Najjar SA, et al. Optogenetic activation of the distal colon epithelium engages enteric nervous system circuits to initiate motility patterns. Am J Physiol-Gastrointest Liver Physiol. 2021;321(4):G426-435.
- Böhn L, et al. Self-reported food-related gastrointestinal symptoms in IBS are common and associated with more severe symptoms and reduced quality of life. Off J Am Coll Gastroenterol. 2013;108(5):634–41.
- Atkinson W, et al. Food elimination based on IgG antibodies in irritable bowel syndrome: a randomised controlled trial. Gut. 2004;53(10):1459–64.
- Pittayanon R, et al. Gut microbiota in patients with irritable bowel syndrome—a systematic review. Gastroenterology. 2019;157(1):97–108.
- Hugerth LW, et al. No distinct microbiome signature of irritable bowel syndrome found in a Swedish random population. Gut. 2020:69(6):1076–84.
- Jeffery IB, et al. An irritable bowel syndrome subtype defined by speciesspecific alterations in faecal microbiota. Gut. 2012;61(7):997–1006.
- Jalanka-Tuovinen J, et al. Faecal microbiota composition and hostmicrobe cross-talk following gastroenteritis and in postinfectious irritable bowel syndrome. Gut. 2014;63(11):1737–45.
- El-Salhy M, Hausken T, Hatlebakk JG. Current status of fecal microbiota transplantation for irritable bowel syndrome. Neurogastroenterol Motil. 2021;33(11):e14157.
- Liu Y, et al. Increased ileal immunoglobulin A production and immunoglobulin A-coated bacteria in diarrhea-predominant irritable bowel syndrome. Clinical and Translational. Gastroenterology. 2020;11(3):e00146.
- Paik CN, et al. The role of small intestinal bacterial overgrowth in postgastrectomy patients. Neurogastroenterol Motil. 2011;23(5):e191–6.
- 80. Pimentel M, et al. ACG Clinical Guideline: small intestinal bacterial overgrowth. Off J Am Coll Gastroenterol. 2020;115(2):165–78.
- Moraru IG, et al. Small intestinal bacterial overgrowth is associated to symptoms in irritable bowel syndrome. Evidence from a multicentre study in Romania. Rom J Intern Med. 2014;52(3):143–50.
- Ghoshal UC, et al. A meta-analysis on small intestinal bacterial overgrowth in patients with different subtypes of irritable bowel syndrome. J Gastroenterol Hepatol. 2020;35(6):922–31.
- Shah A, et al. Small intestinal bacterial overgrowth in irritable bowel syndrome: a systematic review and meta-analysis of case-control studies. Am J Gastroenterol. 2020;115(2):190–201.

- 84. Ghoshal UC, Shukla R, Ghoshal U. Small intestinal bacterial overgrowth and irritable bowel syndrome: a bridge between functional organic dichotomy. Gut and Liver. 2017;11(2):196–208.
- 85. Yao CK, Tuck CJ. The clinical value of breath hydrogen testing. J Gastroenterol Hepatol. 2017;32:20–2.
- Shah A, Morrison M, Holtmann GJ. Gastroduodenal "dysbiosis": a new clinical entity. Curr Treat Opt Gastroenterol. 2018;16(4):591–604.
- 87. Simren M. Use and abuse of hydrogen breath tests. Gut. 2006:55(3):297–303.
- Rezaie A, et al. Hydrogen and methane-based breath testing in gastrointestinal disorders: the North American Consensus. Am J Gastroenterol. 2017:112(5):775–84.
- Sundin OH, et al. Does a glucose-based hydrogen and methane breath test detect bacterial overgrowth in the jejunum? Neurogastroenterol Motil. 2018;30(11):e13350.
- Srivastava D, et al. Associations between IL-1RA polymorphisms and small intestinal bacterial overgrowth among patients with irritable bowel syndrome from India. Neurogastroenterol Motil. 2014;26(10):1408–16.
- 91. Rivkin A, Rybalov S. Update on the management of diarrhea-predominant irritable bowel syndrome: focus on rifaximin and eluxadoline. Pharmacother: J Hum Pharmacol Drug Ther. 2016;36(3):300–16.
- Xu D, et al. Rifaximin alters intestinal bacteria and prevents stress-induced gut inflammation and visceral hyperalgesia in rats. Gastroenterology. 2014;146(2):484–496.e4.
- 93. Wauters L, et al. Duodenal inflammation: an emerging target for functional dyspepsia? Expert Opin Ther Targets. 2020;24(6):511–23.
- 94. Wang C, Fang X. Inflammation and overlap of irritable bowel syndrome and functional dyspepsia. J Neurogastroenterol Motil. 2021;27(2):153–64.
- 95. von Wulffen M, et al. Overlap of irritable bowel syndrome and functional dyspepsia in the clinical setting: prevalence and risk factors. Dig Dis Sci. 2019;64(2):480–6.
- Spiller R. Postinfectious functional dyspepsia and postinfectious irritable bowel syndrome: different symptoms but similar risk factors. Gastroenterology. 2010;138(5):1660–3.
- 97. Talley NJ. What causes functional gastrointestinal disorders? A proposed disease model. Am J Gastroenterol. 2020;115(1):41–8.

# **Publisher's Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

# Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- $\bullet\,$  thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

# At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

